Synthesis of 3'- and 4'-deoxyfluorolactose and its Me₃SiCH₂CH₂ and ceramide † derivatives

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ABSTRACT

(2-Trimethylsilyl)ethyl (Me₃SiCH₂CH₂) 3'- and 4'-deoxyfluorolactosides (1 and 3) were synthesized by glycosylation of Me₃SiCH₂CH₂ 2,3,6-tri-O-benzyl-β-D-glucopyranoside with 2,4,6-tri-O-acetyl-3-deoxy-3-fluoro-β-D-galactopyranosyl bromide and 2,3,6-tri-O-benzyl-4-deoxy-4-fluoro-β-D-galactopyranosyl bromide. Anomeric deblocking of the fully acetyled Me₃SiCH₂CH₂ glycosides (12 and 13) gave the corresponding hemiacetals 14 and 15. Removal of the acetyl groups gave 3'- and 4'-deoxyfluorolactose (2 and 4). The deoxyfluorolactosylceramides 5 and 6 were synthesized via boron trifluoride etherate- or silver triflate-activation of the trichloroacetimidates prepared from 14 and 15. Silver triflate-mediated glycosylations showed lower reaction rates, and fewer byproducts were formed.

INTRODUCTION

Cell-surface glycoconjugates, such as glycolipids and glycoproteins, are important in cell adhesion and immunological responses to carbohydrate antigens^{1,2}. Lactose is a common part of these glycoconjugates. A number of lectins and bacterial adhesins with lactose specificity have been identified recently^{3,4}. Deoxyand deoxyfluorolactosides^{5,6}, are potentially useful tools for investigating cell-adhesion phenomena where lactose-specific proteins are involved. The methyl 6- and 6'-monodeoxyfluoro- β -lactosides were recently reported⁶. We now report the synthesis of the 3'- and 4'-monodeoxyfluorolactose derivatives 1-6.

Simple methyl pyranosides are readily available starting materials for the synthesis of oligosaccharides; however, methyl glycosides are not easily transformed into useful glycoconjugates. In contrast, Me₃SiCH₂CH₂ pyranosides permit conversion, typically in > 90% yield, into the corresponding 1-O-acyl-, hemiacetal, and 1-chloro-sugars, en route to glycoconjugates⁷⁻⁹. Normally, these transfor-

[†] Part 10 of the series 2-(Trimethylsilyl)ethyl (Me₃SiCH₂CH₂) Glycosides for part 9, see ref 17.

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mations are performed with oligosaccharides obtained late in multistep syntheses, and high yields are therefore important. Cleavage of methyl glycosides are in most cases better performed via acetolysis followed by deacetylation by reagents that are selective for 1-acetates¹⁰, although the overall yield from such reaction sequences are normally lower than those obtained by direct Me₃SiCH₂CH₂ cleavage. The latter approach is exemplified here by the synthesis of the 2-deoxyfluorolactosylceramides 5 and 6.

SYNTHESIS

The 3- and 4-deoxyfluoro-galactose derivatives 7 and 8 were prepared as described¹¹ and transformed into 2,4,6-tri-O-acetyl-3-deoxy-3-fluoro- α -D-galactopyranosyl bromide (7a) and 2,3,6-tri-O-benzyl-4-deoxy-4-fluoro- α -D-galactopyranosyl bromide (8a), respectively. Glycosylation of Me₃SiCH₂CH₂ 2,3,6-tri-O-benzyl- β -D-glucopyranoside⁷ (9) with the galactosyl bromides 7a and 8a in the presence of silver triflate and silver silicate, gave the protected deoxyfluorolactosides 10 (75%) and 11 (65%). Hydrogenolytic debenzylation of 10, followed by conventional acetylation, gave the acetylated deoxyfluorolactoside 12 (98%).

Deacetylation of 12 with methanolic sodium methoxide gave the unprotected deoxyfluorolactoside 1 (99%). Hydrogenolytic debenzylation of 11 gave the deoxyfluorolactoside 3 (98%), which was acetylated to give 13 (99%).

The Me₃SiCH₂CH₂ group of 1 and 3 was removed by treatment with trifluoroacetic acid in dichloromethane⁷ to give the 3'- and 4'-deoxyfluorolactoses 2 (95%) and 4 (95%). Compounds 2 and 4 were also prepared by deacetylation of the hemiacetals 14 and 15.

Removal of the $Me_3SiCH_2CH_2$ group in 12 and 13 was performed as above⁷ to give the acetylated deoxyfluoro hemiacetals 14 (95%) and 15 (93%). Treatment of the hemiacetals 14 and 15 with trichloroacetonitrile and DBU^{12} gave the corresponding α -trichloroacetimidates 16 and 17 in essentially quantitative yield after chromatography. $BF_3 \cdot Et_2O$ -catalyzed glycosylation¹² of the azidosphingosine 22 (ref 13) by the donors 16 and 17 afforded the β -glycosides 18 (55%) and 19 (50%), respectively. The corresponding α -glycosides were also isolated in $\sim 15\%$ yield.

Recently, it was suggested¹⁴ that silver triflate would be a more suitable agent for activatation of anomeric thrichloroacetimidates. A slight modification of this method permitted the glycosylation of 22 with trichloroacetimidates 16 and 17, which gave 18 (65%) and 19 (60%). The corresponding α -glycosides were formed in $\sim 5\%$ yield, and the unreacted donors (16 and 17) could, to some extent, be recovered from the reaction mixtures.

The azido groups in 18 and 19 were reduced by hydrogen sulfide 15 , and the primary amino groups were subsequently acylated by octadecanoic acid 16 to give 20 and 21 in > 90% overall yield. Finally, deacylation of 20 and 21 using methanolic sodium methoxide afforded the deoxyfluorolactosylceramides 5 and 6.

EXPERIMENTAL

General methods.—All transformations except hydrogenolyses were performed under Ar. 2,4,6-Tri-O-acetyl-3-deoxy-3-fluoro- α -D-galactopyranosyl bromide (7a) was prepared as previously described 11a. 2,3,6-Tri-O-benzyl-4-deoxy-4-fluoro- α -D-galactopyranosyl bromide (8a) was prepared by treatment of the hemiacetal 8 (ref 11b) with oxalyl bromide and DMF 11c. 1H NMR spectra were recorded with a Varian XL-300 spectrometer. 19F NMR spectra were recorded with external CF₃CO₂H as standard. Chemical shifts (δ , ppm) are relative to internal CFCl₃. Optical rotations were measured with a Perkin–Elmer 141 polarimeter. TLC was performed on Kieselgel 60 F₂₅₄ (Merck) with detection by UV light and/or by charring with H₂SO₄. Column chromatography was performed on Kieselgel 60 (Grace, 35–70 μ m). Powdered molecular sieves were activated by heating with a flame until no more water condensed on the inside of the flask. Chloroform and CH₂Cl₂ were dried by passage through alumina (Merck, neutral, activity grade 1) immediately before use.

2-(Trimethylsilyl)ethyl 4-O-(3-deoxy-3-fluoro- β -D-galactopyranosyl)- β -D-glucopyranoside (1).—Compound 12 (70.0 mg, 0.1 mmol) was dissolved in dry MeOH (1 mL) and a catalytic amount of NaOMe was added. The solution was stirred at room temperature. TLC (9:1 EtOAc-MeOH and 200:150:3 CHCl₃-MeOH-H₂O) showed that 12 was completely consumed after 12 h. The solution was

Scheme 1.

neutralized by addition of silica, filtered (Celite), and concentrated. The residue was dissolved in water and freeze-dried to give 1 (44 mg, 99%); $[\alpha]_D^{25} - 17^\circ$ (c 1, H_2O); ¹H NMR (D_2O): δ 5.00 (d, 1 H, J 7.7 Hz, H-1'), 4.80 (ddd, 1 H, J 49.8, 10.0, 3.2 Hz, H-3'), 4.39 (d, 1 H, J 7.8 Hz, H-1), 4.00 (ddd, 1 H, J 10.0, 7.8, 3.3 Hz, H-2'), 1.00 (ddt, 2 H, CC H_2Si); 0.00 [s, 9 H, Si(C H_3)₃]; ¹⁹F NMR data (D_2O): δ 220 (dd, J 49.5, 3 Hz, F-3').

4-O-(3-Deoxy-3-fluoro-β-D-galactopyranosyl)-D-glucopyranose (2).—(a) Compound 1 (44 mg, 0.10 mmol) was dissolved in CH_2Cl_2 (0.5 mL) and CF_3CO_2H (1 mL) was added. The mixture was stirred for 30 min at room temperature, 1:2 propyl acetate—toluene (9 mL) was added and the solution was concentrated. A second portion of toluene was added and evaporated. Column chromatography of the residue (SiO₂, 100:50:1 CHCl₃-MeOH-H₂O) gave 2 (33 mg, 95%).

(b) Compound 14 (60 mg, 0.1 mmol) was dissolved in dry MeOH (1 mL) and a catalytic amount of NaOMe was added. The solution was stirred overnight at room temperature, neutralized (SiO₂), and concentrated. Column chromatography (SiO₂, 100:50:1 CHCl₃-MeOH-H₂O) of the residue gave 2 (31 mg, 89%, slightly

contaminated with a faster-moving impurity according to TLC, but pure according to NMR); $[\alpha]_D^{25} + 45^\circ$ (2 h at 25°C, c 2, H₂O); ¹H NMR (D₂O): δ 5.10 (d, 1 H, J 7.8 Hz, H-1'), 4.90 (ddd, 1 H, J 49.8, 10.0, 3.2 Hz, H-3'), 5.60, 4.45 (2d, 1 H, J 3.3, 7.8 Hz, H-1 α , H-1 β), 4.20 (ddd, 1 H, J 10.0, 7.8, 3.3 Hz, H-2'); ¹⁹F NMR data (D₂O): δ 220 (dd, J 49.5, 3 Hz, F-3').

2-(Trimethylsilyl)ethyl 4-O-(4-deoxy-4-fluoro-β-D-galactopyranosyl)-β-D-glucopyranoside (3).—Compound 11 (102.0 mg, 0.10 mmol) was dissolved in AcOH (10 mL) and hydrogenolyzed (H₂, 1 atm, 50 mg Pd–C, 10%; room temperature) during 12 h. The mixture was filtered (Celite) and concentrated. Column chromatography of the residue (9:1 EtOAc-MeOH) gave 3 (43.6 mg, 98%); $[\alpha]_D^{25}$ –15° (c 1, H₂O); ¹H NMR (D₂O): δ 4.90 (dd, 1 H, J 51.1, 3.2 Hz, H-4′), 4.85 (d, 1 H, J 7.7 Hz, H-1′), 4.42 (d, 1 H, J 7.9 Hz, H-1), 3.80 (ddd, 1 H, J 29.1, 10.0, 3.2 Hz, H-3′), 0.95 (ddt, 2 H, CCH₂Si), 0.00 [s, 9 H, Si(CH₃)₃]; ¹⁹F NMR data (D₂O): δ 220 (dt, J 51.1 and 30.1 Hz, F-4′).

4-O-(4-Deoxy-4-fluoro-β-D-galactopyranosyl)-D-glucopyranose (4).—(a) Compound 3 (44 mg, 0.10 mmol) was treated with CF_3CO_2H as described for the preparation of 2. Column chromatography (SiO₂, 100:50:1 CHCl₃-MeOH-H₂O) gave 4 (33 mg, 95%).

(b) Compound 15 (60 mg, 0.1 mmol) was treated with methanolic NaOMe as described for 2. Column chromatography gave 4 (32 mg, 92%); $[\alpha]_D^{25} + 47^\circ$ (2 h at 25°C, c 2, H₂O); ¹H NMR (D₂O): δ 5.00 (dd, 1 H, J 51.1, 3.4 Hz, H-4'), 4.85 (d, 1 H, J 7.7 Hz, H-1'), 5.51, 4.42 (2d, 1 H, J 3.1, 7.8 Hz, H-1 α , H-1 β); ¹⁹F NMR (D₂O): δ 220 (dt, J 51.1 and 29.8 Hz, F-4').

4-O-(3-Deoxy-3-fluoro-β-D-galactopyranosyl)-β-D-glucopyranosyl-(1 \rightarrow 1)-(2S, 3R,4E)-2-N-octadecanoylsphingenine (5).—Compound 20 (12.5 mg, 0.01 mmol) was dissolved in MeOH (1 mL) and NaOMe (5 mg, 0.1 mmol) was added. The mixture was stirred for 12 h at room temperature, silica was added and the stirring was continued for 1 h. Filtration of the mixture (Celite), evaporation of the solvent, and column chromatography (9:1 EtOAc-MeOH) of the amorphous residue gave 5 (8.4 mg, 94%); $[\alpha]_D^{25}$ – 5° (c 1, CDCl₃); ¹H NMR (CD₃OD): δ (lactose unit) 4.98 (d, 1 H, J 7.9 Hz, H-1'), 4.81 (ddd, 1 H, J 50.1, 9.9, 3.3 Hz, H-3'), 4.41 (d, 1 H, J 7.9 Hz, H-1); 4.00 (ddd, 1 H, J 10.0, 7.9, 3.5 Hz, H-2'), (ceramide unit) 5.71 (dt, 1 H, J 15.4, 7.6 Hz, C=CHCH₂), 5.48 (dd, 1 H J 15.4, 7.5 Hz, CH=C), 2.05 (m, 2 H, C=CHCH₂), 1.96 (m, 2 H, CH₂CO), 1.41–1.15 (m, 54 H, 27 CH₂), 0.87 (bt, 6 H, J 6.5 Hz, 2 CH₃). Anal. Calcd for C₄₈H₉₀FO₁₂N: C, 64.6; H 10.2. Found: C, 64.7; H, 10.3.

4-O-(4-Deoxy-4-fluoro-β-D-galactopyranosyl)-β-D-glucopyranosyl-(1 \rightarrow 1)-(2S, 3R,4E)-2-N-octadecanoylsphingenine (6).—Treatment of compound 21 (12.6 mg, 0.01 mmol) as described for 5, gave 6 (8.5 mg, 96%); [α]_D²⁵ -8° (c 1, CDCl₃); ¹H NMR (CD₃OD): δ (lactose unit) 4.89 (dd, 1 H, J 50.8, 3.4 Hz, H-4'), 4.87 (d, 1 H, J 8.0 Hz, H-1'), 4.40 (d, 1 H, J 7.9 Hz, H-1), 3.78 (ddd, 1 H, J 29.7, 10.0, 3.3 Hz, H-3'), (ceramide unit) 5.70 (dt, 1 H, J 15.4, 7.6 Hz, C=CHCH₂), 5.48 (dd, 1 H, J 15.4, 7.6 Hz, CH=C), 2.05 (m, 2 H, C=CHCH₂), 1.95 (m, 2 H, CH₂CO), 1.41-1.15

(m, 54 H, 27 CH₂), 0.88 (bt, 6 H, J 6.5 Hz, 2 CH₃). Anal. Calcd for C₄₈H₉₀FO₁₂N: C, 64.6; H 10.2. Found: C, 64.6; H, 10.3.

2-(Trimethylsilyl)ethyl 2,3,6-tri-O-benzyl-4-O-(2,4,6-tri-O-acetyl-3-deoxy-3-fluoro-β-D-galactopyranosyl)-β-D-glucopyranoside (10).—Compound 9 (ref 7, 681 mg, 1.2 mmol), silver trifluoromethanesulfonate (308 mg, 1.2 mmol), and N,N,N',N'-te-tramethylurea (180 μL, 1.5 mmol), were dissolved in dry CH₂Cl₂ (10 mL) and 3A molecular sieves (0.7 g) were added. The mixture was stirred at room temperature for 30 min and then cooled to -20° C. Freshly prepared 2,4,6-tri-O-acetyl-3-deoxy-3-fluoro-α-D-galactopyranosyl bromide (7a)^{11a} (370 mg, 1 mmol) in CH₂Cl₂ (5 mL) was added dropwise during 15 min. The mixture was stirred overnight at room temperature. The mixture was filtered (Celite) and concentrated. Flash chromatography of the residue (1:3 EtOAc-heptane) gave 10 (630 mg, 75%); $[\alpha]_D^{25} - 8^{\circ}$ (c 1, CHCl₃); ¹H NMR (CDCl₃): 5.50 (dd, 1 H, J 4.4, 3.2 Hz, H-4'), 5.30 (ddd, 1 H, J 10.4, 8.1, 3.1 Hz, H-2'), 5.10 (d, 1 H, J 8.1 Hz, H-1'), 4.90 (ddd, 1 H, J 50.0, 10.4, 3.2 Hz, H-3'), 4.95-4.60 (m, 6 H, CHPh), 4.37 (d, 1 H, J 7.7 Hz, H-1), 2.12, 2.01, 1.97 (3 s, 3 H each, OAc), 1.03 (m, 2 H, CCH₂Si), 0.03 [s, 9 H, Si(CH₃)₃]. Anal. Calcd for C₄₄H₅₇FO₁₃Si: C, 62.8; H 6.8. Found: C, 62.8; H, 6.8.

2-(Trimethylsily)ethyl 2,3,6-tri-O-benzyl-4-O-(2,3,6-tri-O-benzyl-4-deoxy-4-fluoro-β-D-galactopyranosyl)-β-D-glucopyranoside (11).—2,3,6-Tri-O-benzyl-4-deoxy-4-fluoro-α-D-galactopyranosyl bromide (8a) (prepared as described 11c from 250 mg, 0.54 mmol of 8, ref 11b) and 9 (ref 7, 363 mg, 0.64 mmol) were dissolved in CH₂Cl₂ (2.5 mL, distilled from CaH₂). Activated 3A molecular sieves (350 mg) were added and the system was stirred under Ar at room temperature for 1 h. Silver silicate (350 mg) was added, the mixture was protected from light, and the stirring was continued at room temperature for 5 h. The mixture was filtered and concentrated and the residue was chromatographed (SiO₂, 2:1 EtOAc-heptane) to give 11 (352 mg, 65% based on 8); $[\alpha]_D^{25} + 8^{\circ}$ (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 4.90 (dd, 1 H, J 51.0, 3.3 Hz, H-4'), 4.84 (d, 1 H, J 7.8 Hz, H-1'), 4.42 (d, 1 H, J 7.9 Hz, H-1), 3.70 (ddd, 1 H, J 29.2, 9.8, 3.4 Hz, H-3'), 1.01 (ddt, 2 H, CCH₂Si), 0.00 [s, 9 H, Si(CH₃)₃]. Anal. Calcd for C₅₉H₆₉FO₁₀Si: C, 71.9; H, 7.1. Found: C, 72.0; H, 7.1.

2-(Trimethylsilyl)ethyl 2,3,6-tri-O-acetyl-4-O-(2,4,6-tri-O-acetyl-3-deoxy-3-fluoro-β-D-galactopyranosyl)-β-D-glucopyranoside (12).—Compound 10 (500 mg, 0.59 mmol) was dissolved in AcOH (30 mL) and hydrogenolyzed (H_2 , 1 atm, 150 mg Pd-C, 10%; room temperature) during 12 h. The mixture was filtered (Celite) and concentrated. The residue was dissolved in 2:7 Ac₂O-pyridine (10 mL), and stirred overnight at room temperature. Concentration of the solution and column chromatography of the residue (1:2 EtOAc-heptane) gave 12 (403 mg, 98%); $[\alpha]_D^{25}$ - 10° (c 1, CHCl₃); ¹H NMR (CDCl₃): 5.70 (dd, 1 H, J 4.0, 3.3 Hz, H-4'), 5.30 (ddd, 1 H, J 10.4, 8.1, 3.1 Hz, H-2'), 5.18 (dd, 1 H, J 9.5, 9.1 Hz, H-3), 4.90 (ddd, 1 H, J 50.0, 10.4, 3.2 Hz, H-3'), 5.10 (d, 1 H, J 8.1 Hz, H-1'), 4.89 (dd, 1 H, J 9.7, 7.8 Hz, H-2), 4.50 (m, H-6 or 6'), 4.03-4.20 (m, 3 H, H-6,6'), 4.37 (d, 1 H, J 7.7 Hz, H-1), 2.20-2.00 (5 s, 18 H, OAc), 1.01 (m, 2 H, CCH₂Si), 0.00 [s, 9 H, Si(CH₃)₃]. Anal. Calcd for C₂₉H₄₅FO₁₆Si: C, 50.0; H 6.5. Found: C, 49.9; H, 6.5.

- 2-(Trimethylsilyl)ethyl 2,3,6-tri-O-acetyl-4-O- (2,3,6-tri-O-acetyl-4-deoxy-4-fluoro-β-D-galactopyranosyl)-β-D-glucopyranoside (13).—Compound 3 (20 mg, 0.045 mmol) was dissolved in 2:1 Ac₂O-pyridine (1 mL). The solution was stirred overnight at room temperature and then concentrated. Column chromatography of the residue (1:2 EtOAc-heptane) gave 13 (31 mg, 99%); $[\alpha]_{5}^{25}$ -8° (c 1, CHCl₃); ¹H NMR (CDCl₃): δ 5.79 (dd, 1 H, J 51.0, 3.1 Hz, H-4'), 5.30 (ddd, 1 H, J 30.0, 10.4, 3.3 Hz, H-3'), 5.25 (dd, 1 H, J 9.4, 9.1 Hz, H-3), 5.21 (dd, 1 H, J 10.3, 7.4 Hz, H-2'), 4.61 (d, 1 H, J 7.9 Hz, H-1'), 4.52 (m, 2 H, H-1,6), 4.12 (dd, 1 H, J 12.1, 5.1 Hz, H-6), 3.80-4.00 (m, 3 H, CHCSi, H-4,5'), 3.66 (m, 1 H, H-5), 3.50 (dt, 1 H, CHCSi), 3.45 (2 H, H-6'), 2.13-1.94 (4 s, 18 H, OAc), 1.02 (m, 2 H, CCH₂Si), 0.01 [s, 9 H, Si(CH₃)₃]. Anal. Calcd for C₂₉H₄₅FO₁₆Si: C, 50.0; H 6.5. Found: C, 50.0; H, 6.5.
- 2,3,6-Tri-O-acetyl-4-O-(2,4,6-tetra-O-acetyl-3-deoxy-3-fluoro-β-D-galactopyranosyl)-D-glucopyranose (14).—Compound 12 (69.7 mg, 0.10 mmol) was treated with CF₃CO₂H as described for the preparation of 2. Column chromatography of the residue (1:1 EtOAc-heptane) gave 14 (57 mg, 95%) as an α/β mixture; ¹H NMR (CDCl₃): 5.60, 4.50 (2d, 1 H, J 3.0 and 7.7 Hz, H-1 α , H-1 β), 4.90 (ddd, 1 H, J 50.0, 10.4, 3.2 Hz, H-3'), 5.00 (d, 1 H, J 8.0 Hz, H-1'), 2.20–2.00 (m, 18 H, OAc). Anal. Calcd for C₂₄H₃₃FO₁₆: C, 48.3; H 5.6. Found: C, 48.3; H, 5.5.
- 2,3,6-Tri-O-acetyl-4-O-(2,3,6-tetra-O-acetyl-4-deoxy-4-fluoro-β-D-galactopyranosyl)-D-glucopyranose (15).—Compound 13 (69.7 mg, 0.10 mmol) was treated with CF₃CO₂H as described for the preparation of 2. Column chromatography (1:1 EtOAc-heptane) gave 15 (55 mg, 93%) as an α/β mixture; ¹H NMR (CDCl₃): δ 5.10 (dd, 1 H, J 51.0, 3.4 Hz, H-4'), 4.90 (d, 1 H, J 7.7, H-1'), 5.56, 4.42 (2d, 1 H, J 3.4 and 7.9 Hz, H-1 α , H-1 β), 2.10–1.90 (m, 18 H, OAc). Anal. Calcd for C₂₄H₃₃FO₁₆: C, 48.3; H 5.6. Found: C, 48.1; H, 5.6.
- 2,3,6-Tri-O-acetyl-4-O-(2,4,6-tetra-O-acetyl-3-deoxy-3-fluoro- β -D-galactopyranosyl)- β -D-glucopyranosyl-(1 \rightarrow 1)-(2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecen-1,3-diol (18).—To a solution of compound 14 (238 mg, 0.4 mmol) in CH₂Cl₂ (4 mL) was added trichloroacetonitrile (201 μ L, 2.0 mmol) and DBU (8 μ L, 0.05 mmol) at 0°C. The mixture was allowed to attain room temperature. The reaction was shown to be complete after 6 h (TLC), where upon the mixture was concentrated. The residue was chromatographed (SiO₂, 1:1 EtOAc-heptane) and the α -trichloroacetimidate 16 [¹H NMR (CDCl₃): δ 8.50 (s, 1 H, CNH), 6.70 (d, 1 H, J 3.4 Hz, H-1)] was divided in two portions of equal size and treated as follows.
- (a) To a solution of 16 (147 mg) and 22 (129 mg, 0.3 mmol) in CH_2Cl_2 (2 mL, containing 4A molecular sieves) at 0°C was added $BF_3 \cdot Et_2O$ (27 μ L, 0.22 mmol). The mixture was kept for 3 h at 0°C. The mixture was diluted with CH_2Cl_2 (5 mL), washed with satd aq NaHCO₃, dried (Na₂SO₄), filtered, and concentrated. Column chromatography (SiO₂, 3:1 \rightarrow 1:1 EtOAc-heptane) gave 18 (111 mg, 55%).
- (b) Compound 16 (147 mg), 22 (129 mg, 0.3 mmol), and silver triflate (57 mg, 0.22 mmol) in a dark, round-bottomed flask was dried on a vacuum pump overnight. The flask was filled with Ar and $\mathrm{CH_2Cl_2}$ (2 mL) was added at room temperature. The reaction was allowed to proceed for 18 h, then the mixture was

treated as for (a) to give **18** (131 mg, 65%); $[\alpha]_D^{25} - 7^\circ$ (c 1, CHCl₃); ¹H NMR (CDCl₃) δ : (lactose unit) 5.69 (dd, 1 H, J 4.0, 3.3 Hz, H-4'), 5.32 (ddd, 1 H, J 10.3, 8.0, 3.2 Hz, H-2'), 5.20 (dd, 1 H, J 9.9, 9.2 Hz, H-3); 4.87 (ddd, 1 H, J 50.0, 10.3, 3.2 Hz, H-3'), 5.07 (d, 1 H, J 7.9 Hz, H-1'); 4.89 (dd, 1 H, J 9.8, 7.8, Hz, H-2), 4.50 (m, H-6 or 6'), 4.05–4.18 (m, 3 H, H-6,6'), 4.35 (d, 1 H, J 7.8 Hz, H-1), 2.18–2.00 (5 s, 18 H, OAc); (azidosphingosine unit) 5.80 (dt, 1 H, J 15.3, 7.6 Hz, H-5), 5.60 (dd, 1 H, J 7.8, 4.8 Hz, H-3); 5.41 (dd, 1 H, J 15.3, 7.6 Hz, H-4), 3.90 (m, CHN₃) 2.04 (bd, 2 H, J 7.5 Hz, H-6), 1.35–1.15 (m, 22 H, CH₂), 0.85 (t, 3 H, J 6.5 Hz, CH₃). Anal. Calcd for C₄₉H₇₀FN₃O₁₈: C, 58.4; H 7.0. Found: C, 58.3; H, 6.9.

2,3,6-Tri-O-acetyl-4-O-(2,3,6-tri-O-acetyl-4-deoxy-4-fluoro-β-D-galactopyranosyl)-β-D-glucopyranosyl-($1 \rightarrow 1$)-(2S,3R,4E)-2-azido-3-O-benzoyl-4-octadecen-1,3-diol (19).—Compound 15 (214 mg, 0.36 mmol) was treated as for 14 to give the α-trichloroacetimidate 17 [¹H NMR (CDCl₃): δ 8.60 (s, 1 H, CNH); 6.60 (d, 1 H, J 3.4 Hz, H-1)], and then 19 (91 mg, 50% yield according to procedure a, and 109 mg, 60% yield according to procedure b); [α] $_D^{25}$ – 5° (c 1, CHCl₃); 1 H NMR (CDCl₃): δ (lactose unit) 5.79 (dd, 1 H, J 51.0, 3.3 Hz, H-4'), 5.31 (ddd, 1 H, J 30.1, 10.4, 3.3 Hz, H-3'), 5.26 (dd, 1 H, J 9.4, 9.0 Hz, H-3), 5.20 (dd, 1 H, J 10.3, 7.7 Hz, H-2'), 4.62 (d, 1 H, J 7.8 Hz, H-1'), 4.49 (m, 2 H, H-1,6), 4.13 (dd, 1 H, J 12.0, 5.2 Hz, H-6), 3.79–3.97 (m, 3 H, H-4,5'), 3.64 (m, 1 H, H-5), 3.42 (2 H, H-6'), 2.08–1.95 (5 s, 18 H, OAc), (azidosphingosine unit) 5.79 (dt, 1 H, J 15.4, 7.6 Hz, H-5), 5.61 (dd, 1 H, J 7.8, 4.8 Hz, H-3), 5.41 (dd, 1 H, J 15.3, 7.6 Hz, H-4), 3.90 (m, CHN₃), 2.01 (m, 2 H, H-6), 1.35–1.15 (m, 22 H, 11 CH₂), 0.87 (t, 3 H, J 6.5 Hz, CH₃). Anal. Calcd for C₄9H₇₀FN₃O₁8: C, 58.4; H 7.0. Found: C, 58.5; H, 7.0.

2,3,6-Tri-O-acetyl-4-O-(2,4,6-tetra-O-acetyl-3-deoxy-3-fluoro-β-D-galactopyranosyl)- β -D-glucopyranosyl- $(1 \rightarrow 1)$ -(2S,3R,4E)-3-O-benzoyl-2-N-octadecanoylsphingenine (20).—Hydrogen sulfide was bubbled through a solution of 18 (50 mg, 0.05 mmol) in 10:1 pyridine-Et₃N (10 mL)¹⁵ for 2 h. The flask was sealed and the solution was stirred for 24 h. Argon was bubbled through the solution to remove H₂S. The solvent was removed and the residue (which was used in the subsequent step without further purification) was dissolved in CH₂Cl₂ (5 mL). Octadecanoic acid (28 mg, 0.1 mmol) and 1-ethyl-3-(3-dimethylaminoisopropyl)carbodiimide hydrochloride (EDC, 29 mg, 0.15 mmol) was added¹⁶ and the mixture was stirred at room temperature for 18 h, then CH₂Cl₂ (15 mL) was added, the solution was washed with water, dried (Na2SO4), and concentrated. The residue was chromatographed (SiO₂, 70:1 CH₂Cl₂-MeOH) to give 20 (59 mg, 95%) ¹H NMR (CDCl₃) δ: (lactose unit) 5.80 (dd, 1 H, J 51.0, 3.2 Hz, H-4'), 5.30 (ddd, 1 H, J 30.0, 10.4, 3.3 Hz, H-3'), 5.27 (dd, 1 H, J 9.4, 9.0 Hz, H-3), 5.19 (dd, 1 H, J 10.3, 7.6 Hz, H-2'), 4.62 (d, 1 H, J 7.8 Hz, H-1'), 4.49 (m, 2 H, H-1,6), 4.13 (dd, 1 H, J 12.0, 5.2 Hz, H-6), 3.79–3.99 (m, 3 H, H-4,5'), 3.64 (m, 1 H, H-5), 3.42 (2 H, H-6'), 2.10-1.95 (4 s, 18 H, OAc), (ceramide unit) 5.68 (dt, 1 H, J 15.3, 7.6 Hz, $C=CHCH_2$), 5.58 (dd, 1 H, J 7.8, 4.8 Hz, CHOBz), 5.48 (dd, 1 H, J 15.3, 7.6 Hz, CH=C), 2.01 (m, 4 H, H-6, CH₂CO), 1.40–1.15 (m, 54 H, CH₂), 0.86 (bt, 6 H, J 6.5 Hz, CH₃). Anal. Calcd for C₆₇H₁₀₆FNO₁₉: C, 64.5; H 8.6. Found: C, 64.4; H, 8.6.

2,3,6-Tri-O-acetyl-4-O-(2,3,6-tri-O-acetyl-4-deoxy-4-fluoro-β-D-galactopyranosyl)-β-D-glucopyranosyl-($1 \rightarrow 1$)-(2S,3R,4E)-3-O-benzoyl-2-N-octadecanoylsphingenine (21).—Compound 19 (50 mg, 0.05 mmol) was reduced with H₂S and treated with octadecanoic acid-EDC as for 18 to give 21 (58 mg, 91%); [α]_D²⁵ – 7° (c 1, CHCl₃); ¹H NMR (CDCl₃) δ: (lactose unit) 5.79 (dd, 1 H, J 51.0, 3.1 Hz, H-4′), 5.30 (ddd, 1 H, J 30.0, 10.4, 3.3 Hz, H-3′), 5.25 (dd, 1 H, J 9.4, 9.1 Hz, H-3), 5.21 (dd, 1 H, J 10.3, 7.4 Hz, H-2′), 4.61 (d, 1 H, J 7.9 Hz, H-1′), 4.52 (m, 2 H, H-1,6), 4.12 (dd, 1 H, J 12.1, 5.1 Hz, H-6), 3.80–4.00 (m, 3 H, H-4,5′), 3.66 (m, 1 H, H-5), 3.45 (2 H, H-6′), 2.13–1.94 (4 s, 18 H, OAc), (ceramide unit) 5.67 (dt, 1 H, J 15.3, 7.6 Hz, C=CHCH₂), 5.58 (dd, 1 H, J 7.7, 4.8 Hz, CHOBz), 5.48 (dd, 1 H, J 15.4, 7.6 Hz, CH=C), 2.01 (m, 4 H, C=CHCH₂, CH₂CO), 1.41–1.15 (m, 54 H, 27 CH₂), 0.86 (bt, 6 H, J 6.5 Hz, CH₃). Anal. Calcd for C₆₇H₁₀₆FNO₁₉: C, 64.5; H 8.6. Found: C, 64.4; H, 8.5.

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REFERENCES

- 1 S. Hakomori, in S. Sell (Ed.), Serological Cancer Markers, The Humana Press, Totowa, NL, 1992, pp. 207-232.
- 2 (a) N. Sharon and H. Lis, Science, 246 (1989) 227-234; (b) J.L. Winkelhake, Glycoconjugate J., 8 (1991) 381-386.
- 3 (a) K.-A. Karlsson, Annu. Rev. Biochem., 58 (1989) 309-350; (b) V. Jimenez-Lucho, V. Ginsburg and H.C. Krivan, Infect. Immun., 58 (1990) 2085-2090; (c) K. Saukkonen, W.N. Burnette, V.L. Mar, H.R. Masure and E.I. Tuomanen, Proc. Natl. Acad. Sci. USA, 89 (1992) 118-122.
- 4 H. Leffler, F.R. Masiarz and S.H. Barondes, Biochemistry., 28 (1989) 9222-9229.
- 5 (a) K. Adelhorst and K. Bock, Carbohydr. Res., 202 (1990) 131-149; (b) A. Rivera-Sagredo, J. Jiminez-Barbero, M. Martin-Lomas, D. Solis and T. Diaz-Mauriño, Carbohydr. Res., 232 (1992) 207-226; (c) T. Ekberg and G. Magnusson, Carbohydr. Res., in press.
- 6 S. Cai, S. Hakomori and T. Toyokuni, J. Org. Chem., 57 (1992) 3431-3437.
- 7 K. Jansson, S. Ahlfors, J. Dahmén, T. Frejd, J. Kihlberg, G. Magnusson, G. Noori and K. Stenvall, J. Org. Chem., 53 (1988) 5629-5647.
- 8 K. Jansson, G. Noori and G. Magnusson, J. Org. Chem., 55 (1990) 3181-3185.
- 9 G. Magnusson, Trends Glycosci. Glycotechnol., 4 (1992) 358-367.
- 10 N.V. Bovin, S.E. Zurabayan and A.Y. Khorlin, J. Carbohydr. Chem., 2 (1983) 249-262.
- (a) P. Kovac and P.J. Glaudemans, Carbohydr. Res., 123 (1983) 326-331; (b) J. Kihlberg, T. Frejd, K. Jansson, S. Kitzing and G. Magnusson, Carbohydr. Res., 185 (1989) 171-190; (c) C.A.A. van Boeckel, T. Beetz, S.F. van Aelst, Tetrahedron, 40 (1984) 4097-4107.
- 12 R.R. Schmidt, Pure Appl. Chem., 61 (1989) 1257-1270; P. Sinaÿ, ibid., 63 (1991) 519-528.
- 13 P. Zimmerman and R.R. Schmidt, Liebigs Ann. Chem., (1988) 663-667.
- 14 S.P. Douglas, D.M. Whitfield and J.J. Krepinsky, J. Carbohydr. Chem., 12 (1993) 131-136.
- 15 J. Kihlberg and D. Bundle, Carbohydr. Res., 216 (1991) 65-78.
- 16 T. Murase, H. Ishida, M. Kiso and A. Hasegawa, Carbohydr. Res., 188 (1989) 71-80.
- 17 U. Ellervik and G. Magnusson, Acta Chem. Scand., in press.